

# Progressive methods for investigation of migration of pharmaceuticals in soils

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## Introduction

the migration of pharmaceuticals in soils = **important role in toxicity and bioavailability**  
**knowledge of transport behaviour** essential for the prediction of their biotoxicity  
 soil "self-cleaning" ability related to the **organic matter** = immobilization = stable complexes  
 decrease in concentration of free mobile drugs = **decrease in ability to enter into environment**

## Experimental

**humic acids (HA)** = representant of organic matter  
**sulphapyridine (SPY)** = representant of antibiotics  
 diffusion experiments *in situ* and model hydrogels  
 agarose hydrogel enriched by HA and HA hydrogels

$c$  = concentration  
 $c_s$  = concentration at interface  
 $m_t$  = total diffusion flux in time  $t$   
 $x$  = distance from interface  
 $D$  = diffusion coefficient  
 $L$  = thickness of hadrogel layer  
 $\varepsilon$  = partition coefficient  
 $S$  = area  
 $n$  = SPY pulse at interface

Eq. (1)  $m_t = 2 \cdot c_s \cdot \sqrt{\frac{D \cdot t}{\pi}}$       Eq. (2)  $c = c_s \cdot \operatorname{erfc} \frac{x}{2 \cdot \sqrt{D \cdot t}}$

Eq. (3)  $\vec{j} = -D \cdot \varepsilon \cdot \frac{\Delta c}{L}$

Eq. (4)  $c = \frac{n}{S \cdot \sqrt{\pi \cdot D \cdot t}} \cdot \exp\left(-\frac{x^2}{4 \cdot D \cdot t}\right)$       Eq. (5)  $\ln c = \ln \frac{n}{S \cdot \sqrt{\pi \cdot D \cdot t}} - \frac{x^2}{4 \cdot D \cdot t}$

**diffusion from constant source**  
 cuvette filled by hydrogel immersed in saturated solution  
**diffusion couple**  
 donor SPY hydrogel connected with pure acceptor hydrogel

**diffusion cells**  
 hydrogel between donor and acceptor compartments

**diffusion from instantaneous planar source**  
 filter paper saturated by SPY as source for hydrogel  
***in situ***  
 soil column in real conditions (filter paper ...)

## Results & Discussion

Results obtained for agarose hydrogel allows to differentiate between the effects of porous structure of hydrogel and interactions between HA and SPY.

The  $D$  values obtained for pure inert hydrogel:

$(1.01 \pm 0.05) \times 10^{-10} \text{ m}^2 \text{ s}^{-1}$   
(diffusion couple);

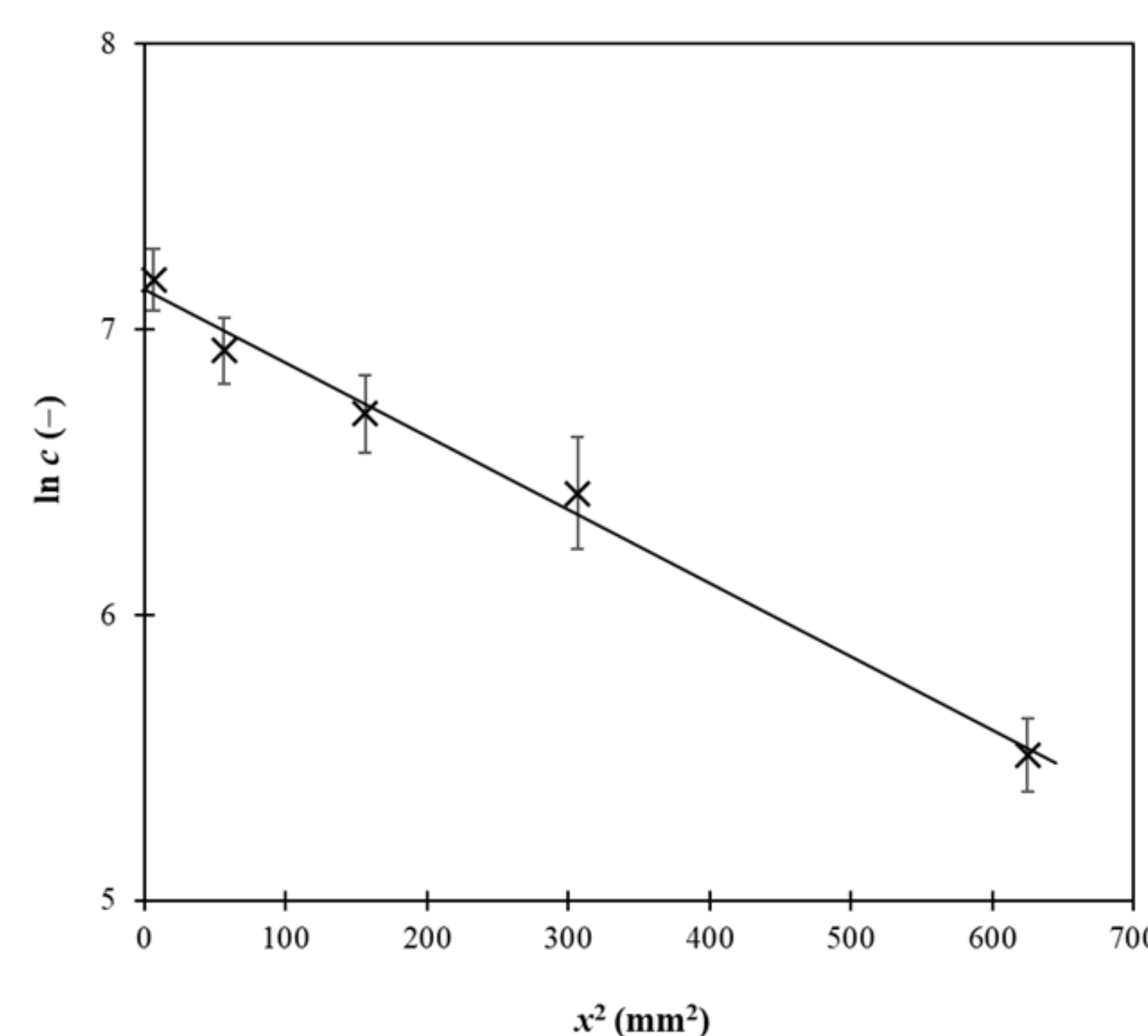
$(4.60 \pm 0.21) \times 10^{-10} \text{ m}^2 \text{ s}^{-1}$   
(constant source);

$(4.93 \pm 0.38) \times 10^{-10} \text{ m}^2 \text{ s}^{-1}$   
(diffusion cells).

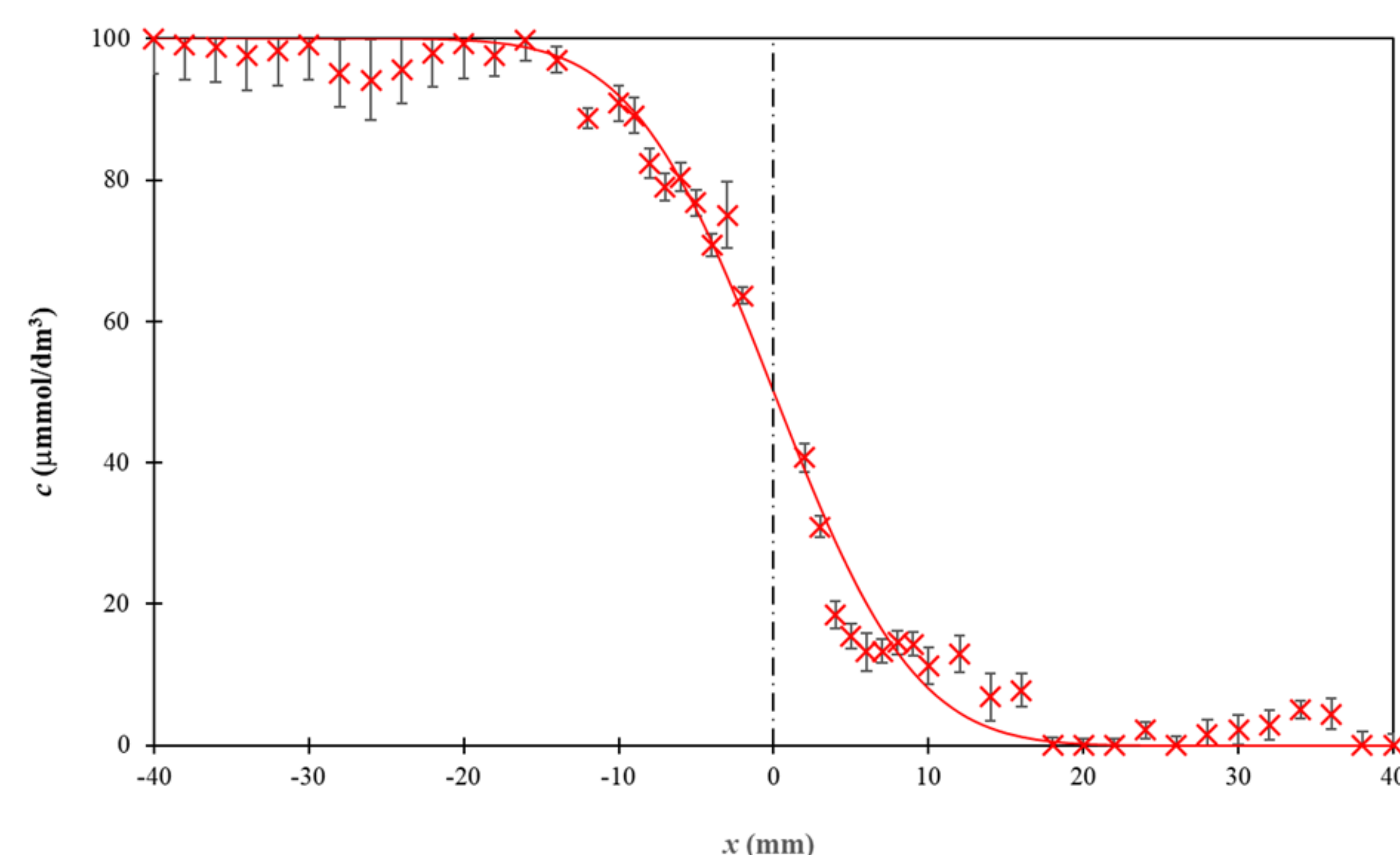
Comparing with  $D$  value ( $2.03 \times 10^{-9} \text{ m}^2 \text{ s}^{-1}$ ) for the diffusion of SPY in aqueous solution (10.1016/j.arabjc.2013.12.009), the decrease caused by the pore structure is observed.

The HA addition in porous hydrogel resulted in the additional decrease in  $D$  value caused by the interaction between SPY and HA and immobilization of SPY in hydrogel structure.

matrix	method	diffusion coefficient ( $\text{m}^2 \text{ s}^{-1}$ )
enriched agarose hydrogel	diffusion couple	$(9.85 \pm 0.47) \times 10^{-11}$
	constant source	$(3.40 \pm 0.13) \times 10^{-10}$
	diffusion cells	$(4.49 \pm 0.12) \times 10^{-10}$
humic hydrogel soil	instantaneous planar source	$(4.38 \pm 0.29) \times 10^{-10}$
	<i>in situ</i>	$(4.23 \pm 0.27) \times 10^{-10}$



The example of experimental data fitting using Eq. (5).



The example of experimental profiles for diffusion couple after 72 h fitted by Eq. (2).

## Conclusions

Both model mediums (agarose hydrogel enriched by HA and HA hydrogel) can provide practically the same results as *in situ* measurements in soil, if a suitable experimental method is chosen. This finding is very usable for a prediction of the behaviour of SPY and pharmaceuticals (and other pollutants) in soil environments.